

Mesenchymal Stem Cells in the Treatment of Amyotrophic Lateral Sclerosis

Mahsa Hajivalili^{1,2}, Fatemeh Pourgholi^{1,2}, Hossein Samadi Kafil³, Farhad Jadidi-Niaragh^{1,2,4} and Mehdi Yousefi^{1,2,3,*}

¹Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran; ²Department of Immunology, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran; ³Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran; ⁴Department of Immunology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran



Mehdi Yousefi

Abstract: Amyotrophic lateral sclerosis (ALS) is a neurodegenerative disorder which is characterized by motor neuron (MN) dysfunction, progressive paralysis, and death. Although several therapeutic approaches have been used for treatment of ALS, little success has been achieved. Natural vectors such as mesenchymal stem cells (MSCs) can be a promising tool for overcoming therapeutic problems. MSCs have multipotential characteristics such as the ability to differentiate into variety of cell types, easy access, immunomodulation, tissue repair, exertion of trophic factors, exosome secretion and efficient homing. In this review, we will discuss the characteristics of MSCs and their possible therapeutic mechanisms in ALS patients.

Keywords: Amyotrophic lateral sclerosis, mesenchymal stem cells, stem cell, treatment.

1. INTRODUCTION

Amyotrophic lateral sclerosis (ALS) is a rare fatal neurodegenerative disease which is characterized by the loss of upper and lower motor neurons resulting in muscle weakness, progressive paralysis and mortality 3 to 5 years after the clinical onset due to respiratory failure [1]. It is estimated that about 30,000 people in United States are struggling with ALS [2] with a higher incidence rate in men [3]. ALS can be divided into two forms, including sporadic ALS (sALS) with 90% prevalence and the familial ALS (fALS) affecting only 10% of patients with a possible link to some genetic mutations [4]. Up to now 13 genes have been introduced that can affect ALS inheritance. *SOD1* (encodes for copper/zinc ion-binding superoxide dismutase), *TARDBP* (also known as *TDP-43*; encodes for TAR DNA binding protein), *OPTN* (encodes optineurin), *ANG* (encodes angiogenin, ribonuclease, RNase A family, 5) and *FUS* (encodes fusion in sarcoma) were clinically important. Mutated *SOD1* is the leading cause for 20% of familial ALS, although 5-10% of familial ALS mutations are related to *TARDBP*, mutations in *FUS* stands for 5%, and mutations in *ANG* is found in around 1% of ALS patients [5].

The presence of blood brain barrier (BBB) is the main problem in the treatment of central nervous system (CNS)-related diseases such as ALS [6]. The newly designated methods especially drug delivery vectors may help to solve this problem. There are two types of drug delivery tools,

including synthetic and natural vectors. In general, synthetic vectors are products of chemical reactions from lipophilic compounds. Liposomes are the main type of synthetic tools utilized by researchers, however, regarding their possible side effects such as immunogenicity, accumulation in tissues and unknown consequences of this method, applying natural vectors instead is being considered [7].

Bacteria, viruses and stem cells are the most frequent vectors which have been used as drug delivery tools, however, application of stem cells is more interesting. Stem cell therapy as a branch of regenerative medicine is a method of employing natural vectors as therapeutic drug delivery systems [8, 9]. Among the cellular vectors that have been used for treatment of medical conditions, embryonic stem cells (ESCs) and adult stem cells (ASCs) are more popular. Regarding the ethical issues of ESCs, their employment is usually associated with limitations [10-12]. Therefore, ASCs are more attractive for clinical research. Adult stem cells (ASCs) especially mesenchymal stem cells (MSCs) have several features which make them an ideal option for drug delivery purpose. MSCs are multipotential [13] tissue resident cells [14] which have high capacity of differentiation into specialized cells especially mesodermal-derived types, however differentiation to other lineages is also possible [15, 16]. MSCs exhibit their therapeutic functions through different mechanisms such as tissue repair [17], immunomodulation [18], exertion of trophic factors [19] and efficient homing [20].

As there is no definite cure for ALS, transplantation of naive or engineered MSCs (EMSCs) may be considered as novel promising approach for treatment of ALS. There is evidence which shows EMSCs can control the disease pro-

*Address correspondence to this author at the Department of Immunology, School of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran; Tel: +98-4133364665; Fax: +98-4133364665; E-mail: Yousefime@tbzmed.ac.ir

gression by several mechanisms in ALS animal models [21]. In this review, we will discuss about the new advances in biology of MSCs and their role in the treatment of ALS.

2. MSCs CHARACTERISTICS

The Friedenstein's group described the unique characteristics of MSCs for the first time, which has been further completed by Owen and coworkers [22]. MSCs are fibroblast-like, multipotent progenitors which have self-renewal capacity and high differentiation rate [23]. They usually differentiate into mesodermal-derived cells, however, they can also generate other cell lineages including endoderm and ectoderm-derived ones [15, 24-26]. Bio-preservation of MSCs is possible and during preservation they revealed minimum loss of potency [27]. It has been reported that MSCs are present in every vascularized tissues [28]. They can be easily isolated from bone marrow, adipose tissue, umbilical cord, derm, pancreas, liver, muscles and lungs. Moreover, under specific conditions, expansion of MSCs will be possible [29-32].

3. MSC ISOLATION

Due to lack of consensus marker for purification of MSCs, the International Society for Cellular Therapy (ISCT) has introduced minimum criteria containing adhesion to plastic surfaces and positive and/or negative expression of some markers. It has been demonstrated that MSCs can express molecules such as CD73, CD90 and CD105. On the other hand, CD34, CD45, CD19 and HLA class II molecules do not express on MSCs [33, 34]. In addition, presence of different surface receptors will be helpful for their isolation. Recently, the application of monoclonal antibodies for isolation of MSCs has become popular. One of the most useful monoclonal antibodies for this purpose is against Stro-1 [35]. Several markers beside Stro-1 have been proposed for isolation of MSCs, including CD146 (MUC-18), CD271 (low affinity nerve growth factor) [35] and the embryonic stem cell marker (SSEA-4) [36]. Other properties related to MSCs such as phenotypic difference [37, 38], required growth factors [39], chemokine [40, 41] and cytokine [42] receptors, cell-matrix and cell-cell receptors [43] may also be helpful for purification of different MSCs from different organs [35]. As the *in vitro* expansion of MSCs is associated with lower genetic abnormalities compared to other stem cell types, it seems that the rate of malignancy induction by MSC therapy is slight [44].

4. MSCs ORIGIN

Due to lack of discriminating markers, isolation of MSC progenitors is very difficult. Little is known regarding the genetic mechanisms behind the lineage differentiation management in MSC progenitors. Identification of these mechanisms may lead to the production of MSCs from ESCs [45].

As bone marrow aspiration method is very invasive, investigation for identification of new sources for isolation of MSCs is going on [46]. As mentioned previously wide range of organs are suitable for MSCs separation, however, umbilical cord blood [47], placental tissue [48] and adipose tissue [49] have been considered as potent substitutions of BM-MSCs. Despite the existence of phenotypic similarities be-

tween MSCs derived from different organs, they differ in function which is in part due to organ specific niche [47-49].

5. THERAPEUTIC MECHANISMS OF MSCs

As mentioned previously, the therapeutic mechanisms of MSCs are in part through the excretion of trophic factors, immunomodulation, anti-inflammatory effects, efficient homing and exosome secretion. Cooperation of these therapeutic mechanisms will lead to tissue repair and disease attenuation. Here, we will discuss each mechanism.

5.1. Generation of Trophic Factors

MSCs can generate several trophic factors and natural proteins which are crucial for neuronal survival. Interaction of MSCs with TLR ligands, inflammatory cytokines and conditions such as hypoxia are the main stimulators of various trophic factors secretion [50]. The majority of trophic factors are mediators of angiogenesis and apoptosis inhibitors [51, 52]. Among the MSCs-derived growth factors, brain-derived neurotrophic factor (BDNF) [53], ciliary neurotrophic factor (CNTF) [54], glial derived neurotrophic factor (GDNF) [55], nerve growth factor (NGF) [56], insulin-like growth factor1 (IGF-1) [57], and vascular endothelial growth factor (VEGF) play an important role in neurons replenishment and protection [58]. Transferring of growth factors which are usually large peptides to the CNS faced with an BBB obstacle [59]. It has been shown that genetic-modified stem cells generate neurotrophic and growth factors which are essential for MN regeneration through different mechanisms such as clearance of toxic compounds and production of glial cells [60].

5.2. Immunomodulation

It has generally been accepted that MSCs have potent immunomodulatory functions [61]. It has been demonstrated that MSCs can shift immune response from TH1 (cell mediated) toward TH2 (humoral) [62]. It has also been suggested that MSCs may inhibit the production of CD4⁺ T cell cytokines such as IL-2 and IFN- γ . The induced anergy in T cells following coculture with MSCs is in part due to the lack of costimulatory molecules on MSCs. This coculture may even lead to generation of suppressor T cells [63, 64].

The controversial results have been reported about the effect of MSCs on B cells. Suppression of B cell proliferation and stimulation of antibody secretion are some of MSCs effects on B cells [65]. It has been suggested that MSCs inhibit NK cells cytotoxic activity through down regulation NKp30 and NKG2 receptors [66].

The hMSCs exude several immunomodulatory factors such as IDO (Indoleamine 2,3-dioxygenase), Semaphorin-3A, B7-H4, HLA-G, LIF (Lukemia Inhibitory Factor), TSG6 (Tumor necrosis factor-inducible gene 6 protein) and Galectins. These immunosuppressing factors can inhibit inflammatory process and induce T regulatory cells, reversibly [67].

5.3. Anti-Inflammatory Effects

The anti-inflammatory function of MSCs is in part due to up regulation of TGF- β . MSCs can also inhibit inflammatory

process through reduction of oxidative stress and peroxide dismutase secretion [68]. MSCs engraftment on the third day after spinal cord injury resulted in shifting of macrophage phenotype from M1 toward M2 [68]. M1 macrophage belongs to pro-inflammatory group whereas M2 phenotype defined as anti-inflammatory ones.

5.4. Efficient Homing

There are several problems regarding tracking of MSCs homing *in vivo*, including lack of discriminating markers and low concentration of MSCs in blood stream [69]. It has been shown that MSCs can infiltrate into different injured tissues such as ischemic brain injury [70], myocardial infarction [71] and acute renal failure [72]. Although expression of several chemokine receptors including CCR1, CCR4, CCR7, CCR9, CCR10, CXCR1, CXCR3, CXCR4, CXCR5 and CX3CR1 has been observed [40, 41], the predominant chemokines in MSCs homing are not recognized yet. Among the chemokines responsible for leukocytes homing, stromal derived factor -1 (SDF-1) has been studied more precisely [73]. It has been shown that SDF-1 is the main mediator of stromal progenitor cells migration into injured tissues in a rat model of myocardial infarction [74]. Abbott and colleagues showed that SDF-1 was over expressed in ischemic muscles following myocardial infarction, which enhanced the mobilization of endothelial progenitor cells [75]. Subsequently, Ip *et al.* exhibited that $\beta 1$ integrins (but not CXCR4) are the main molecules in MSCs homing [76].

The efficient homing of MSCs is under control of three molecular groups including chemokines (particularly CXCR4, CXCL12, CCL/R2), adhesion molecules (such as integrins), and matrix metallo proteases (especially MMP-2) [77, 78]. The transplantation of CXCR4 transfected-MSCs into rat model of myocardial infarction has been associated with ameliorative effects [79]. Furthermore, *in vitro* and *in vivo* analysis of MSCs revealed that P-selectin, vascular cell adhesion protein-1 (VCAM-1) and very late antigen-4 (VLA-4) are the most important factors for interaction of MSCs with endothelial cells [80]. Surprisingly, injection of VCAM-1 transfected MSCs in contrast with naive MSCs enhanced homing into inflammatory sites in a mouse model of inflammatory bowel disease [81]. This experiment implicates a manipulation possibility of homing molecules in order to improve MSCs homing ability which can lead to better therapeutic outcomes. Furthermore, efficient homing can be affected by the route of MSC administration and origin of cells, for example intravenously administered MSCs mostly entrapped in lungs and showed little migration to the CNS [82]. Eliopoulos *et al.* demonstrated that allogenic MSCs could be successfully administrated without considering of immune recognition and rejection by host immune system [83].

The cell confluence and number of passages are two most important factors in culture of MSCs. It has been shown that increased confluency in cultured MSCs leads to secretion of inhibitory compounds like TIMP-3 (a natural MMP-inhibitor) and dysregulated trans-endothelial migration following injection. However, *in vitro* expanded MSCs in contrast with freshly isolated MSCs, do not express some critical receptors for homing, particularly CXCR4. Moreover, this

loss can be compensated by cytokine cocktail in MSCs culture medium [69]. Choi *et al.* showed that approximately after ten passages, the ability of MSCs for neuroprotection, immunomodulation and differentiation has been decreased significantly, so which it seems that the use of freshly isolated MSCs must be a priority [84].

5.5. Exosome Secretion

One of the newly discovered aspects of MSCs is their high capacity of exosome secretion. Exosomes can be defined as secreted membrane vesicles with the ability to carry cargos such as proteins and RNAs for intercellular communications [85]. Tian sheng chen *et al.* have been shown that MSCs derived exosomes were enriched in pre-miRNAs. Their team detected 106 miRNA in MSCs [86]. MSCs exosomes has gained widespread attention not only for their specific composition but also for their successfully therapeutic outcomes in animal models and immunosuppressive activity. Yu B *et al.* reported that transferring exosomes derived from MSC overexpressing GATA-4 (MSC(GATA-4)) upon transplantation in injured cardiomyocytes of rat heart contributed to tissue protection by secreting different miRs, particularly miR-19a, working as inducers of cell survival signaling pathway [86]. In another study Amarnath S *et al.* showed that in a human-into-mouse xenogeneic graft-versus-host disease (x-GVHD) model, mediated by human CD4+ Th1 cells, BMSCs inverted experimental x-GVHD through marked inhibition of Th1 cell effector functions. Serum CD73 expressing exosomes were detected in BMMSC recipients which promoted adenosine accumulation immunosuppression [87]. In a recent study, transplanted MSCs in rats after traumatic brain injury (TBI) revealed functional recovery and neurovascular remodeling through endogenous angiogenesis and neurogenesis by reduction of inflammation process [88]. Nakamura Y *et al.* exhibited that MSC-derived exosomes were able to enhance myogenesis and angiogenesis and induce muscle regeneration, by miRNAs such as miR-494. Therefore MSC derived exosomes could be a useful cell-free therapy tool for different types of neurological disorders [89].

6. ROUTES OF MSCs DELIVERY

There are two major drug delivery methods including remote delivery and direct delivery, which have their own advantages and disadvantages. The intramuscular, intraneural and intravascular delivery routes are examples of remote delivery. Although privileged options as noninvasiveness seems quite enough for efficiency of this method, high dosage of therapeutic agents is required. However direct injection and intrathecal delivery provide the chance to distribute greater therapeutic agents, they are highly invasive [90].

7. ALS AND CURRENT TREATMENTS

Obvious genetic linkage to point mutations in cytosolic $\text{Cu}^{2+}/\text{Zn}^{2+}$ superoxide dismutase1 (SOD1) has been reported in minority of fALS. Several transgenic animal models which have mutations in different genes such as SOD1, fused in sarcoma (FUS), TAR DNA binding protein (TARDBP) and angiogenine have been generated for fALS [91, 92]. Unfortunately the exact etiology of sALS remains

elusive [5]. Destruction of MNs is complex and it is probably under control of diverse pathways. These pathways may lead to protein aggregation, axonal transport deficiency, changes in calcium concentration, impairment of mitochondrial functions and consequently cellular death [93, 94]. Current treatments for ALS are usually based on maintaining quality of life. Riluzole, baclofen, tizanidine, non-steroidal anti-inflammatory drugs (NSAIDs), tramadol, physical/occupational and speech therapy, and anti-depressant medications are current treatments involved in ALS therapy [95]. All mentioned drugs have some side effects such as dizziness, elevated liver enzymes, granulocytopenia and weakness [96]. Unfortunately, none of the treatments applied for ALS patients are effective and riluzole (the only specific approved drug), is just useful for relieving symptoms and increase patients survival up to two or three months [97]. It seems that the occurrence of hundreds mutations in dozens of genes is involved in etiology of ALS, so which investigators should focus on more genes rather than SOD1. Thus, necessity of approaching to new methods is clear. Development of stem cell therapy as a branch of regenerative medicine, particularly MSCs application is growing rapidly in order to achieve appropriate treatment for ALS [97].

8. MSCs IN THE TREATMENT OF ALS

Recently rodents with G93A-SOD mutations showed conscription of endogenous neural progenitors in deteriorating lumbar spinal cord which was correlated with progression of ALS. This finding provides hopes for creating new therapeutic approach based on stem cells in order to decrease the disease progression. Consistently, it has recently been shown that intravenous, intrathecal, intracerebral and intraspinal administration of MSCs to G93A-SOD1 mice led to advanced motor function, decreased loss of MNs and long time survival [98, 99]. Moreover, intraspinal administration of MSCs to ALS mouse model was decreased the microglial activation [98]. Interestingly, the intravenous injected MSCs to ALS mice could effectively differentiate to neural lineage (as was neurogenin 1 positive), and improve motor function and delay disease onset [100]. Forostyak *et al.* demonstrated that administration of hMSCs in SOD1 rats was associated with prolonged survival, remodeled gene expression paradigm, neurotrophic effects and immunomodulation [101].

The intracerebroventricular administration of glucagon-like peptide1 (peptide with antioxidant functions)-transfected MSCs to G93A-SOD1 mice led to delayed disease onset, generation of astrocyte, microglial activation, decreased neuroinflammation and prolonged survival [102]. Moreover, intramuscular injection of GDNF-transfected MSCs into G93A-SOD1 rats showed improved survival and healthy motor functions. The neuroprotective features of MSCs have been shown by evaluating their ability in secretion of glial-derived neurotrophic factor (GDNF), the vital factor which conserve MNs after brain injury [103]. Although differentiation of MSCs leads to responsiveness of cells to riluzole and upregulation of GDNF, however BM-MSCs in hSOD^{G93A} with the same interventions showed moderate aspartate uptake and unresponsiveness to riluzole therapy [104]. Moreover, Krakora and colleagues showed that application of h-GDNF (human-GDNF) and h-VEGF (human-VEGF) synergistically increased life span [105].

The intraspinal administration of autologous MSCs into 11 Spanish ALS patients was associated with high rate of MN generation [106]. Moreover, it has been demonstrated that intraspinal administration of autologous MSCs was safe and no sign of toxicity or deformity has been showed by two years [107, 108]. Furthermore, there was no report regarding the toxicity or possible tumor generation in the similar studies performed in Israel, India, South Korea and Mexico [109-112] (Fig. 1).

A number of studies have been emphasized on systemic behavior of ALS with effects both on MSC function and motor neurons [84, 113, 114]. Consistently, it has recently been reported that BM-MSCs of ALS patients and healthy individuals could be different in several features despite of their identical cell surface markers and morphology. So which, analysis of Nanog, Oct-4 and Nestin-1 as accepted markers of pluripotency, revealed low expression of Nanog and Oct-4 in ALS MSCs (A-MSCs) [115]. In another study, comparison of ALS hMSCs with hMSCs showed the decreased expression of cytoplasmic FMR interacting protein 2 (CyFIP2) and retinoblastoma (Rb) binding protein 9 (RbBP9) in ALS hMSCs led to dysregulation of their translation [116]. Koh *et al.* have suggested that β -PIX, an intracellular factor involved in migration, is responsible for impaired migration of ALS-hMSCs compared to hMSCs [117]. These findings put some discrepancies in application of autologous stem cell therapy in ALS patients which suggests further investigations.

The main problem regarding the application of MSCs in ALS patients is their low differentiation rate to MNs. One solution for this problem is genetic engineering of MSCs. Consistently, it has been shown that the genetically engineered hMSCs for expression of MNs specific transcription factors such as Olig2 and Hb9, expressed higher levels of MN markers (30% of total cells) and was able to create connections with muscular fibers [118]. In another study, hypoxia-cultured human adipose-derived MSCs (hAMSC-H) applied in GBM showed no sign of tumor forming cooperated with improved viability, tropism and motility toward tumorigenic cells [119].

Kwon *et al.* reported that intrathecal injected MSCs in ALS patients exerted their action by recruitment of immune cells into CNS which was associated with shifting infiltrated cells to T regulatory and TH2 subpopulation. Secretion of IL-4, IL-10 and TGF- β by these cells is one of the indirect immunomodulatory effects of MSC therapy in ALS patients [120].

Regarding the decreased levels of TIMP in ALS patients, pathological upregulation of MMPs may be predictable. As mentioned previously, MMPs are one of the important molecules for MSCs homing into injured MNs. The administration of autologous MSC to patients may associate with three main outcomes including constitution of lost MNs, activation of endogenous neurogenesis and neuroprotective or neurotrophic effects [121].

It has recently been reported that MSCs-derived CX3CL1 can shift activated glial cells toward neuroprotective phenotype [122].

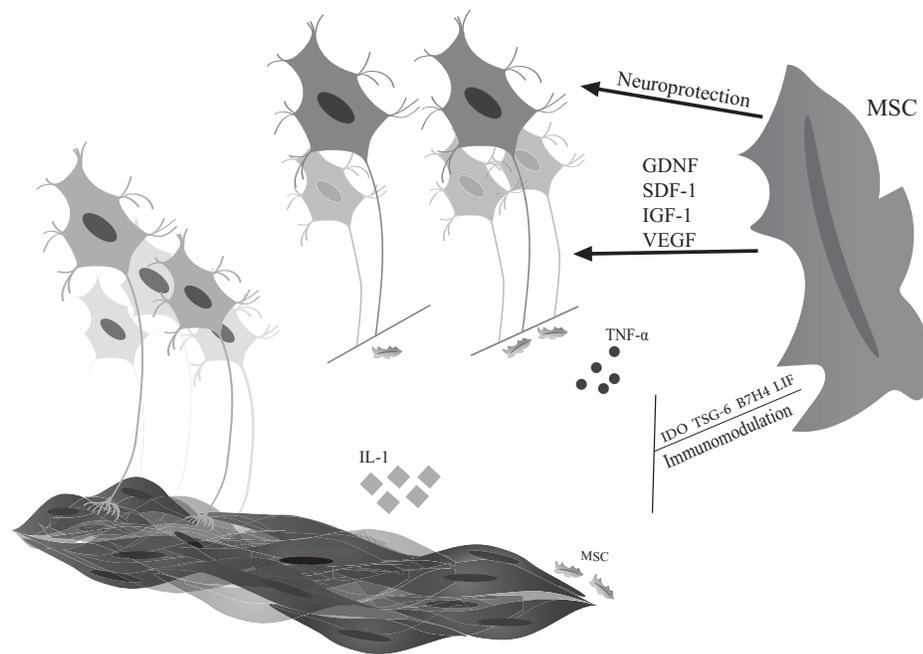


Fig. (1). During onset of ALS and progressive loss of neuromuscular connections, releasing of inflammatory cytokines such as TNF- α and IL-1 leads to recall of MSCs. Neuroprotective effects of migrated MSCs on degenerated neurons is managed by trophic factor secretion where as Immunomodulation aspects are presented by inflammation suppressors.

Table 1. Studies related to application of MSC therapy in mouse model of ALS.

Species	MSC type	Interventions	Results	Reference
G93A-SOD1 murine	hBM-MSCs	Intravenous Intrathecal Intracerebral Intraspinal	Advanced motor function, alleviated loss of MNs, long time survival	[98, 99]
mdf/ocd murine	BM-MSCs	BM-MSCs intraspinal injection	GDNF secretion Neuroprotective effects	[103]
G93A-SOD1 murine	BM-MSCs	Autologous grafts	Moderate aspartate uptake Unresponsiveness to Riluzole	[104]
G93A-SOD1 murine	hBM-MSCs	Intraspinal	Moderating neuroinflammation and activation of microglial cells	[98]
G93A-SOD1 murine	hBM-MSCs expressing neurogenin-1	Intravenous	excellent homing to CNS, improvement in motor function and delayed onset of disease	[100]
SOD1rats	hBM-MSCs	Intrathecal	Rearrangement of gene expression, immunomodulation, neurotrophic effects prolong survival	[101]
G93A-SOD1 murine	Engineered hBM-MSCs expressing GLP-1	Intra cerebro ventricular	generation of astrocyte, microglial activation, decreased rate of neuro inflammation and prolonged survival	[102]
C57BL/6J mice	BM-MSCs	Analyzing MSCs and microglial interactions	Interaction of MSCs CX3CLand microglial CX3CR phenotypic and functional changes of microglia to neuro protective type	[122]
Sprague-Dawley rats	hBM-MSCs	Engineering hMSCs for high expression of olig-2 and Hb-9	Significant neuron features Ability to create connection with muscular fibers	[118]

SOD1: super oxide dismutase1, hBM-MSC: human Bone Marrow- Mesenchymal Stem Cell, MN: Motor Neuron, BM-MSC: Bone Marrow MSC, mdf/ocd: muscle-deficient osteochondrodystrophy, GDNF: Glial Derived Neurotrophic Factor, CNS: Central Nervous System GLP1: Glucagon-Like Peptide 1, olig-2: oligodendrocyte transcription factor gene, Hb-9: home box gene.

Table 2. Studies related to application of MSC therapy in ALS patients.

Species	MSC type	Interventions	Results	Reference
Human	BM-MSCs	Analysis of cells surface markers in ALS patients and healthy individuals	Identical surface markers and morphology Low expression of NANOG, Oct-4, Nestin-1 in ALS MSCs	[115]
Human	BM-MSCs	intrathecally intravenously	Extraordinary rise in CD4 CD25 T regulatory in peripheral blood	[120]
Human	hBM-MSCs	Intraspinal Intrathecal Intraventricular Intravenous	No sign of toxicity and possible tumor forming Prolong survival	[107, 108]
Human	Curcumin nanoparticle loaded hAMSCs	Intravenous	Low cytotoxicity	[123]

MSC: Mesenchymal Stem Cell, ALS: Amyotrophic Lateral Sclerosis, BM-MSC: Bone Marrow MSC, NANOG: transcription factor encoded by NANOG gene, Oct-4: Octamer-binding transcription factor4, hBM-MSC: human BM-MSC, hAMSC: human Adipose MSC.

Table 3. The underway clinical trials related to MSC-based ALS therapy (clinicaltrial.org).

Trial code	Route of injection	Intervention	Status	Country	phase
NCT01609283	Intraspinal	Autologous MSC	Recruiting	USA	I
NCT01142856	Intraspinal	AMSC	completed	USA	I
NCT01494480	Intrathecal	UCMSC	Enrolling by invitation	China	II
NCT01759784	Intraventricular	BMMSC	Not yet recruiting	Iran	I
NCT01759797	Intravenous	BMMSC	completed	Iran	I
NCT02017912	Intramuscular/ Intrathecal	BM MSC-NTF	Recruiting	USA	II
NCT02290886	Intravenous	AMSC	Recruiting	Spain	I/II
NCT01777646	Intramuscular	BMMSC-NTF	Active / not recruiting	Israel	II
NCT01051882	Intramuscular	BM MSC-NTF	completed	Israel	I/II

MSC: Mesenchymal Stem Cell, ALS: Amyotrophic Lateral Sclerosis, AMSC: Adipose MSC, UCMSC: Umbilical Cord MSC, BMMSC: Bone Marrow MSC, BMMSC-NTF: Bone Marrow MSC secreting Neurotrophic Factors.

Application of MSCs in combination with nanoparticles is another promising approach in the treatment of ALS. Tripodo *et al.* reported that targeted drug delivery of incorporated hydrophobic therapeutic agents in a systemic manner via MSCs is a new drug delivery approach which is named carrier in carrier method. The combination therapy with Inulin-d-alfa-tocopherol succinate micelles (INVITE M) and curcumin-loaded MSCs revealed low cytotoxicity compared to delivery of uncoated curcumin by MSCs [123] (Tables 1, 2).

9. MSC BASED THERAPY: TRANSLATION TO CLINIC

The several clinical trials regarding the use of MSCs transplantation in the treatment of different diseases have been registered at www.clinicaltrial.org. Myocardial infarction, graft versus host disease and diabetes are the main diseases that MSC therapy has been used for them.

There are about 30 trials related the CNS diseases such as Alzheimer, multiple sclerosis, Parkinson, glioblastoma, stroke and ALS. Recently 11 studies registered for ALS treatment by different types of MSCs, however most of them hired BM-MSC, UC-MSC and A-MSC [124] (Table 3).

In all of the studies which have been done to evaluate MSCs possible therapeutic mechanisms, there are a number of questions which still remains unanswered and need further investigations. Among the struggling questions we can name the right origin of MSCs, exact route of delivery, best cell population and possible risk of oncogenesis, which have to be considered for each type of diseases.

The best cell numbers injection for optimal responsiveness in human disorders has not been calculated, yet. Optimal cell dosage must be considered as critical issue because of possible toxicity or tolerance mechanisms [125].

Allogenicity of MSCs didn't show any adverse effects after transplantation for tissue repair, however use of fresh MSCs is preferred because *in vitro* expansion of MSCs increases the expression of five major histocompatibility complex class II genes [126, 127].

It has been shown that *in vitro* expansion of MSCs in order to reach adequate number of cells for therapeutic application can be led to malignant transformation of these cells. Consistently, Rubio *et al.* and Wang *et al.* achieved to deliberating data. Former group demonstrated that upregulation of c-myc and downregulation of p16 in human AMSCs will lead to transformation of cultured cells after 4-5 months [128]. However, in later experiment, cultured hBM-MSCs revealed the capability of tumor forming in NOD/SCID mice due to increased activation of telomerase, translocation and chromosome aneuploidism [129]. Regarding these controversial results, the possibility of oncogenesis by MSC based stem cell therapy remains as a conflicting field of study which requires further investigations. As mentioned previously, one of the essential terms of clinical application of MSC therapy is administration of adequate number of cell contents. Based on adverse consequences of increased passage rates, researchers generate MSCs with human telomerase reverse transcriptase gene (hTERT) which provides maintenance of their stemness aspects. Unfortunately genetic abnormalities is the main expecting side effect in this method which probably can become a leading way to oncogenesis [130]. Finally, making definite decision for this challenging item of MSC stem cell therapy needs further investigations.

CONCLUSION

In general the efficacy of MSC therapy in ALS patients is a matter of controversy. The application of MSCs in ALS patients has no ethical issues along with having benefits such as easy access, low risk of immunogenicity and multiple cell sources. Little is known regarding the therapeutic mechanisms exerted by MSCs in treatment of ALS. Improvement of MSC therapeutic functions through MSC engineering is novel promising approach in preclinical studies involved in ALS therapy. It seems that efforts have to be focused on creating the second generation MSCs with optimized activity in comparison with naive MSCs. Application of MSCs in clinic still needs further authorizations from governmental agencies about standard protocols for cell expansion, quality controls for safety issues and introduction of clinical monitoring parameters. We look forward to use MSCs products instead of whole cell injection in order to achieve an easy method of MSCs administration with high safety.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

ACKNOWLEDGEMENTS

Declared none.

REFERENCES

- Chìò A, Mora G, Calvo A, *et al.* Epidemiology of ALS in Italy A 10-year prospective population-based study. *Neurology* 2009; 72(8): 725-31.
- Beghi E, Logroscino G, Chìò A, *et al.* The epidemiology of ALS and the role of population-based registries. *Biochim Biophys Acta* 2006; 1762(11-12): 1150-7.
- Logroscino G, Traynor BJ, Hardiman O, *et al.* Incidence of amyotrophic lateral sclerosis in Europe. *J Neurol Neurosurg Psychiatry* 2010; 81(4): 385-90.
- Howland DS, Liu J, She Y, *et al.* Focal loss of the glutamate transporter EAAT2 in a transgenic rat model of SOD1 mutant-mediated amyotrophic lateral sclerosis (ALS). *Proc Natl Acad Sci USA* 2002; 99(3): 1604-9.
- Kiernan MC, Vucic S, Cheah BC, *et al.* Amyotrophic lateral sclerosis. *Lancet* 2011; 377 (9769): 942-55.
- Rosenberg GA. Neurological diseases in relation to the blood-brain barrier. *J Cereb Blood Flow Metab* 2012; 32(7): 1139-51.
- Torchilin VP. Recent advances with liposomes as pharmaceutical carriers. *Nat Rev Drug Discov* 2005; 4(2): 145-60.
- Yoo J-W, Irvine DJ, Discher DE, Mitragotri S. Bio-inspired, bio-engineered and biomimetic drug delivery carriers. *Nat Rev Drug Discov* 2011; 10(7): 521-35.
- Swi Chang TM. 50th anniversary of artificial cells: their role in biotechnology, nanomedicine, regenerative medicine, blood substitutes, bioencapsulation, cell/stem cell therapy and nanorobotics. *Artif Cells Nanomed Biotechnol* 2007; 35(6): 545-54.
- Leist M, Bremer S, Brundin P, *et al.* The biological and ethical basis of the use of human embryonic stem cells for *in vitro* test systems or cell therapy. *AlteX* 2008; 25(3): 163-90.
- Hentze H, Soong PL, Wang ST, *et al.* Teratoma formation by human embryonic stem cells: evaluation of essential parameters for future safety studies. *Stem cell Res* 2009; 2(3): 198-210.
- Rong Z, Wang M, Hu Z, *et al.* An effective approach to prevent immune rejection of human ESC-derived allografts. *Cell stem cell* 2014; 14(1): 121-30.
- Salem HK, Thiemermann. Mesenchymal stromal cells: current understanding and clinical status. *Stem cells* 2010; 28(3): 585-96.
- Brandau S, Jakob M, Hemeda H, *et al.* Tissue-resident mesenchymal stem cells attract peripheral blood neutrophils and enhance their inflammatory activity in response to microbial challenge. *J Leukoc Biol* 2010; 88(5): 1005-15.
- Ladak A, Olson J, Tredget E, Gordon T. Differentiation of mesenchymal stem cells to support peripheral nerve regeneration in a rat model. *Exp Neurol* 2011; 228(2): 242-52.
- Rodrigo SF, van Ramshorst J, Hoogslag GE, *et al.* Intramyocardial injection of autologous bone marrow-derived *ex vivo* expanded mesenchymal stem cells in acute myocardial infarction patients is feasible and safe up to 5 years of follow-up. *J Cardiovasc Transl Res* 2013; 6(5): 816-25.
- Shin L, Peterson DASHin L, Peterson DA. Human mesenchymal stem cell grafts enhance normal and impaired wound healing by recruiting existing endogenous tissue stem/progenitor cells. *Stem Cells Transl Med* 2013; 2(1): 33-42.
- English K. Mechanisms of mesenchymal stromal cell immunomodulation. *Immunol Cell Biol* 2013; 91(1): 19-26.
- Marconi S, Castiglione G, Turano E, *et al.* Human adipose-derived mesenchymal stem cells systemically injected promote peripheral nerve regeneration in the mouse model of sciatic crush. *Tissue Eng Part A* 2012; 18(11-12): 1264-72.
- Kang SK, Shin IS, Ko MS, Jo JY, Ra JC. Journey of mesenchymal stem cells for homing: strategies to enhance efficacy and safety of stem cell therapy. *Stem Cells Int* 2012; 2012: 342968.
- Phillips LH. Stem cell therapy in ALS: Possible benefit? *Muscle Nerve* 2014; 49(3): 311-2.
- Owen M, Friedenstein A. Stromal stem cells: marrow-derived osteogenic precursors. *Ciba Found Symp* 1988; 136: 42-60.
- Weissman IL. Stem cells: units of development, units of regeneration, and units in evolution. *Cell* 2000; 100(1): 157-68.
- Arthur A, Rychkov G, Shi S, Koblar SA, Gronthos. Adult human dental pulp stem cells differentiate toward functionally active neurons under appropriate environmental cues. *Stem Cells* 2008; 26(7): 1787-95.
- Makino S, Fukuda K, Miyoshi S, *et al.* Cardiomyocytes can be generated from marrow stromal cells *in vitro*. *J Clin Invest* 1999; 103(5): 697-705.
- Zhang P, He X, Liu K, *et al.* Bone marrow stromal cells differentiated into functional Schwann cells in injured rats sciatic nerve. *Artif Cells, Blood Substit Immobil and Biotechnol* 2004; 32(4): 509-18.

- [27] Ho A, Wagner W, Franke W. Heterogeneity of mesenchymal stromal cell preparations. *Cytotherapy* 2008; 10(4): 320-30.
- [28] da Silva Meirelles L, Fontes AM, Covas DT, Caplan. Mechanisms involved in the therapeutic properties of mesenchymal stem cells. *Cytokine Growth Factor Rev* 2009; 20(5-6): 419-27.
- [29] Bianco P, Robey PG, Simmons. Mesenchymal stem cells: revisiting history, concepts, and assays. *Cell Stem Cell* 2008; 2(4): 313-9.
- [30] Pittenger MF, Mackay AM, Beck SC, et al. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999; 284(5411): 143-7.
- [31] Anjos-Afonso F, Bonnet. Nonhematopoietic/endothelial SSEA-1+ cells define the most primitive progenitors in the adult murine bone marrow mesenchymal compartment. *Blood* 2007; 109(3): 1298-306.
- [32] Zuk PA, Zhu M, Mizuno H, et al. Multilineage cells from human adipose tissue: implications for cell-based therapies. *Tissue Eng* 2001; 7(2): 211-28.
- [33] Dominici M, Le Blanc K, Mueller I, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 2006; 8(4): 315-7.
- [34] Porada CD, Almeida-Porada G. Mesenchymal stem cells as therapeutics and vehicles for gene and drug delivery. *Adv Drug Deliv Rev* 2010; 62(12): 1156-66.
- [35] Lv F-J, Tuan RS, Cheung K, Leung VY. Concise Review: The Surface Markers and Identity of Human Mesenchymal Stem Cells. *Stem Cells* 2014; 32(6): 1408-19.
- [36] Gang EJ, Bosnakovski D, Figueiredo CA, Visser JW, Perlingeiro RC. SSEA-4 identifies mesenchymal stem cells from bone marrow. *Blood* 2007; 109(4): 1743-51.
- [37] Docheva D, Padula D, Popov C, et al. Researching into the cellular shape, volume and elasticity of mesenchymal stem cells, osteoblasts and osteosarcoma cells by atomic force microscopy. *J Cell Mol Med* 2008; 12(2): 537-52.
- [38] Delorme B, Chateauvieux S, Charbord P. The concept of mesenchymal stem cells. *Regen Med* 2006; 1(4): 497-509.
- [39] Leo AJ, Grande DA. Mesenchymal stem cells in tissue engineering. *Cells Tissues Organs* 2006; 183(3): 112-22.
- [40] Ringe J, Strassburg S, Neumann K, et al. Towards in situ tissue repair: human mesenchymal stem cells express chemokine receptors CXCR1, CXCR2 and CCR2, and migrate upon stimulation with CXCL8 but not CCL2. *J Cell Biochem* 2007; 101(1): 135-46.
- [41] Chamberlain G, Fox J, Ashton B, Middleton. Concise review: mesenchymal stem cells: their phenotype, differentiation capacity, immunological features, and potential for homing. *Stem Cells* 2007; 25(11): 2739-49.
- [42] Minguell JJ, Erices A, Conget P. Mesenchymal stem cells. *Exp Biol Med* 2001; 226(6): 507-20.
- [43] Docheva D, Popov C, Mutschler W, Schieker M. Human mesenchymal stem cells in contact with their environment: surface characteristics and the integrin system. *J Cell Mol Med* 2007; 11(1): 21-38.
- [44] Wang Y, Zhang Z, Chi Y, et al. Long-term cultured mesenchymal stem cells frequently develop genomic mutations but do not undergo malignant transformation. *Cell Death Dis* 2013; 4(12): e950.
- [45] Gregory CA, Ylostalo J, Prockop DJ. Adult bone marrow stem/progenitor cells (MSCs) are preconditioned by microenvironmental "niches" in culture: a two-stage hypothesis for regulation of MSC fate. *Sci STKE* 2005; 2005(294): pe37.
- [46] Bühring HJ, Treml S, Cerabona F, et al. Phenotypic characterization of distinct human bone marrow-derived MSC subsets. *Ann N Y Acad Sci* 2009; 1176(1): 124-34.
- [47] Romanov YA, Svintsitskaya VA, Smirnov VN. Searching for alternative sources of postnatal human mesenchymal stem cells: candidate MSC-like cells from umbilical cord. *Stem Cells* 2003; 21(1): 105-10.
- [48] Barlow S, Brooke G, Chatterjee K, et al. Comparison of human placenta- and bone marrow-derived multipotent mesenchymal stem cells. *Stem Cells Dev* 2008; 17(6): 1095-107.
- [49] Noël D, Caton D, Roche S, et al. Cell specific differences between human adipose-derived and mesenchymal-stromal cells despite similar differentiation potentials. *Exp Cell Res* 2008; 314(7): 1575-84.
- [50] Cristostomo PR, Wang Y, Markel TA, et al. Human mesenchymal stem cells stimulated by TNF- α , LPS, or hypoxia produce growth factors by an NF κ B-but not JNK-dependent mechanism. *Am J Physiol-Cell Physiol* 2008; 294(3): C675-C82.
- [51] Caplan AI, Dennis JE. Mesenchymal stem cells as trophic mediators. *J Cell Biochem* 2006; 98(5): 1076-84.
- [52] Liu ZC, Chang TM. Preliminary study on intrasplenic implantation of artificial cell bioencapsulated stem cells to increase the survival of 90% hepatectomized rats. *Art Cells, Blood Substit Immobil Biotechnology* 2009; 37(1): 53-5.
- [53] van Velthoven CT, Sheldon RA, Kavelaars A, et al. Mesenchymal stem cell transplantation attenuates brain injury after neonatal stroke. *Stroke* 2013; 44(5): 1426-32.
- [54] Kassis I, Vaknin-Dembinsky A, Karussis D. Bone marrow mesenchymal stem cells: agents of immunomodulation and neuroprotection. *Curr Stem Cell Res Ther* 2011; 6(1): 63-8.
- [55] Gothelf Y, Abramov N, Harel A, Offen D. Safety of repeated transplantations of neurotrophic factors-secreting human mesenchymal stromal stem cells. *Clin Transl Med* 2014; 3(1): 21.
- [56] Blais M, Lévesque P, Bellenfant S, Berthod F. Nerve growth factor, brain-derived neurotrophic factor, neurotrophin-3 and glial-derived neurotrophic factor enhance angiogenesis in a tissue-engineered *in vitro* model. *Tissue Eng Part A* 2013; 19(15-16): 1655-64.
- [57] Ohlson MR. The search for a treatment: researching the use of mesenchymal stem cells that produce insulin-like growth factor 1 as a treatment for amyotrophic lateral sclerosis. 2011-05-11T15: 18: 53Z
- [58] Dai N, Li D, Shi Q, et al. Influence of vascular endothelial growth factor on endothelial components in human bone marrow and umbilical cord mesenchymal stem cells. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 2012; 20(3): 717-21.
- [59] Zhu J, Jiang Y, Xu G, Liu X. Intranasal administration: a potential solution for cross-BBB delivering neurotrophic factors. *Histol Histopathol* 2012; 27(5): 537-48.
- [60] Lladó J, Haenggeli C, Maragakis NJ, Snyder EY, Rothstein JD. Neural stem cells protect against glutamate-induced excitotoxicity and promote survival of injured motor neurons through the secretion of neurotrophic factors. *Mol Cell Neurosci* 2004; 27(3): 322-31.
- [61] Han Z, Jing Y, Zhang S, et al. The role of immunosuppression of mesenchymal stem cells in tissue repair and tumor growth. *Cell Biosci* 2012; 2(1): 8.
- [62] Batten P, Sarathchandra P, Antoniw JW, et al. Human mesenchymal stem cells induce T cell anergy and downregulate T cell alloresponses via the TH2 pathway: relevance to tissue engineering human heart valves. *Tissue Eng* 2006; 12(8): 2263-73.
- [63] Duffy MM, Ritter T, Ceredig R, Griffin MD. Mesenchymal stem cell effects on T-cell effector pathways. *Stem Cell Res Ther* 2011; 2(4): 34.
- [64] Tse WT, Pendleton JD, Beyer WM, Egalka MC, Guinan EC. Suppression of allogeneic T-cell proliferation by human marrow stromal cells: implications in transplantation. *Transplantation* 2003; 75(3): 389-97.
- [65] Corcione A, Benvenuto F, Ferretti E, et al. Human mesenchymal stem cells modulate B-cell functions. *Blood* 2006; 107(1): 367-72.
- [66] Poggi A, Prevosto C, Massaro A-M, et al. Interaction between human NK cells and bone marrow stromal cells induces NK cell triggering: role of NKp30 and NKG2D receptors. *J Immunol* 2005; 175(10): 6352-60.
- [67] Shi Y, Su J, Roberts AI, et al. How mesenchymal stem cells interact with tissue immune responses. *Trends Immunol* 2012; 33(3): 136-43.
- [68] Hawrylyuk GW, Mothe A, Wang J, et al. An *in vivo* characterization of trophic factor production following neural precursor cell or bone marrow stromal cell transplantation for spinal cord injury. *Stem Cells Development* 2011; 21(12): 2222-38.
- [69] Karp JM, Leng Teo GS. Mesenchymal stem cell homing: the devil is in the details. *Cell Stem Cell* 2009; 4(3): 206-16.
- [70] Lee JS, Hong JM, Moon GJ, et al. A Long-Term Follow-Up Study of Intravenous Autologous Mesenchymal Stem Cell Transplantation in Patients With Ischemic Stroke. *Stem Cells* 2010; 28(6): 1099-106.
- [71] Wu Y, Zhao RC. The role of chemokines in mesenchymal stem cell homing to myocardium. *Stem Cell Rev* 2012; 8(1): 243-50.
- [72] Wise AF, Williams TM, Kiewit MB, et al. Human mesenchymal stem cells alter macrophage phenotype and promote regeneration

- via homing to the kidney following ischemia-reperfusion injury. *Am J Physiol-Renal Physiol* 2014; 306(10): F1222-F35.
- [73] Lau TT, Wang D-A. Stromal cell-derived factor-1 (SDF-1): homing factor for engineered regenerative medicine. *Expert Opin Biol Ther* 2011; 11(2): 189-97.
- [74] Yu J, Li M, Qu Z, *et al.* SDF-1/CXCR4-mediated migration of transplanted bone marrow stromal cells toward areas of heart myocardial infarction through activation of PI3K/Akt. *J Cardiovasc Pharmacol* 2010; 55(5): 496-505.
- [75] Zohlh fer D, Ott I, Mehilli J, *et al.* Stem cell mobilization by granulocyte colony-stimulating factor in patients with acute myocardial infarction: a randomized controlled trial. *Jama* 2006; 295(9): 1003-10.
- [76] Ip JE, Wu Y, Huang J, *et al.* Mesenchymal stem cells use integrin β 1 not CXC chemokine receptor 4 for myocardial migration and engraftment. *Mol Biol Cell* 2007; 18(8): 2873-82.
- [77] Wynn RF, Hart CA, Corradi-Perini C, *et al.* A small proportion of mesenchymal stem cells strongly expresses functionally active CXCR4 receptor capable of promoting migration to bone marrow. *Blood* 2004; 104(9): 2643-5.
- [78] Belema-Bedada F, Uchida S, Martire A, Kostin S, Braun T. Efficient homing of multipotent adult mesenchymal stem cells depends on FROUNT-mediated clustering of CCR2. *Cell Stem Cell* 2008; 2(6): 566-75.
- [79] Cheng Z, Ou L, Zhou X, *et al.* Targeted migration of mesenchymal stem cells modified with CXCR4 gene to infarcted myocardium improves cardiac performance. *Mol Ther* 2008; 16(3): 571-9.
- [80] R ster B, G ttig S, Ludwig RJ, *et al.* Mesenchymal stem cells display coordinated rolling and adhesion behavior on endothelial cells. *Blood* 2006; 108(12): 3938-44.
- [81] Ko IK, Kim B-G, Awadallah A, *et al.* Targeting improves MSC treatment of inflammatory bowel disease. *Mol Ther* 2010; 18(7): 1365-72.
- [82] Fischer UM, Harting MT, Jimenez F, *et al.* Pulmonary passage is a major obstacle for intravenous stem cell delivery: the pulmonary first-pass effect. *Stem Cells Dev* 2009; 18(5): 683-92.
- [83] Eliopoulos N, Stagg J, Lejeune L, Pommey S, Galipeau J. Allogeneic marrow stromal cells are immune rejected by MHC class I- and class II-mismatched recipient mice. *Blood* 2005; 106(13): 4057-65.
- [84] Choi MR, Kim HY, Park J-Y, *et al.* Selection of optimal passage of bone marrow-derived mesenchymal stem cells for stem cell therapy in patients with amyotrophic lateral sclerosis. *Neurosci Lett* 2010; 472(2): 94-8.
- [85] Yeo RWY, Lai RC, Zhang B, *et al.* Mesenchymal stem cell: an efficient mass producer of exosomes for drug delivery. *Adv Drug Deliv Rev* 2013; 65(3): 336-41.
- [86] Chen TS, Lai RC, Lee MM, *et al.* Mesenchymal stem cell secretes microparticles enriched in pre-microRNAs. *Nucleic acids Res* 2010; 38(1): 215-24.
- [87] Yu B, Kim HW, Gong M, *et al.* Exosomes secreted from GATA-4 overexpressing mesenchymal stem cells serve as a reservoir of anti-apoptotic microRNAs for cardioprotection. *Int J Cardiol* 2015; 182: 349-60.
- [88] Amarnath S, Foley JE, Farthing DE, *et al.* Bone Marrow Derived Mesenchymal Stromal Cells Harness Purinergic Signaling to Tolerize Human Th1 Cells *In Vivo*. *Stem Cells* 2015; 33(4): 1200-12.
- [89] Zhang Y, Chopp M, Meng Y, *et al.* Effect of exosomes derived from multipotent mesenchymal stromal cells on functional recovery and neurovascular plasticity in rats after traumatic brain injury. *J Neurosurg* 2015; 122(4): 856-67.
- [90] O'Connor DM, Boulis NM. Cellular and molecular approaches to motor neuron therapy in amyotrophic lateral sclerosis and spinal muscular atrophy. *Neurosci Lett* 2012; 527(2): 78-84.
- [91] Maruyama H, Morino H, Ito H, *et al.* Mutations of optineurin in amyotrophic lateral sclerosis. *Nature* 2010; 465(7295): 223-6.
- [92] Beleza-Meireles A, Al-Chalabi A. Genetic studies of amyotrophic lateral sclerosis: controversies and perspectives. *Amyotroph Lateral Scler* 2009; 10(1): 1-14.
- [93] Manfredi G, Xu Z. Mitochondrial dysfunction and its role in motor neuron degeneration in ALS. *Mitochondrion* 2005; 5(2): 77-87.
- [94] Celsi F, Pizzo P, Brini M, *et al.* Mitochondria, calcium and cell death: a deadly triad in neurodegeneration. *Biochim Biophys Acta* 2009; 1787(5): 335-44.
- [95] Majmudar S, Wu J, Paganoni S. Rehabilitation in amyotrophic lateral sclerosis: Why it matters. *Muscle Nerve* 2014; 50(1): 4-13.
- [96] Naganska E, Matyja E. Amyotrophic lateral sclerosis—looking for pathogenesis and effective therapy. *Folia Neuropathol* 2011; 49(1): 1-13.
- [97] Bensimon G, Lacomblez L, Meininger V. A controlled trial of riluzole in amyotrophic lateral sclerosis. *New Engl J Med* 1994; 330(9): 585-91.
- [98] Vercelli A, Mereuta O, Garbossa D, *et al.* Human mesenchymal stem cell transplantation extends survival, improves motor performance and decreases neuroinflammation in mouse model of amyotrophic lateral sclerosis. *Neurobiol Dis* 2008; 31(3): 395-405.
- [99] Mazzini L, Fagioli F, Boccaletti R, *et al.* Mazzini L, Fagioli F, Boccaletti R, *et al.* Stem cell therapy in amyotrophic lateral sclerosis: a methodological approach in humans. *Amyotroph Lateral Scler* 2003; 4(3): 158-61.
- [100] Chan-II C, Young-Don L, Heejaung K, *et al.* Neural induction with neurogenin 1 enhances the therapeutic potential of mesenchymal stem cells in an amyotrophic lateral sclerosis mouse model. *Cell Transplant* 2012; 22(5): 855-70.
- [101] Forostyak S, Homola A, Turnovcova K, *et al.* Intrathecal Delivery of Mesenchymal Stromal Cells Protects the Structure of Altered Perineuronal Nets in SOD1 Rats and Amends the Course of ALS. *Stem Cells* 2014; 32(12): 3163-72.
- [102] Knippenberg S, Thau N, Dengler R, Brinker T, Petri S. Intracerebroventricular injection of encapsulated human mesenchymal cells producing glucagon-like peptide 1 prolongs survival in a mouse model of ALS. *PLoS one* 2012; 7(6): e36857.
- [103] Suzuki M, McHugh J, Tork C, *et al.* Direct muscle delivery of GDNF with human mesenchymal stem cells improves motor neuron survival and function in a rat model of familial ALS. *Mol Ther* 2008; 16(12): 2002-10.
- [104] Boucherie C, Caumont A-S, Maloteaux J-M, Hermans E. *In vitro* evidence for impaired neuroprotective capacities of adult mesenchymal stem cells derived from a rat model of familial amyotrophic lateral sclerosis (hSOD1^{G93A}). *Exp Neurol* 2008; 212(2): 557-61.
- [105] Krakora D, Mulcrone P, Meyer M, *et al.* Synergistic effects of GDNF and VEGF on lifespan and disease progression in a familial ALS rat model. *Mol Ther* 2013; 21(8): 1602-10.
- [106] Blanquer M, Moraleda JM, Iniesta F, *et al.* Neurotrophic bone marrow cellular nests prevent spinal motoneuron degeneration in amyotrophic lateral sclerosis patients: a pilot safety study. *Stem Cells* 2012; 30(6): 1277-85.
- [107] Mazzini L, Mareschi K, Ferrero I, *et al.* Mesenchymal stromal cell transplantation in amyotrophic lateral sclerosis: a long-term safety study. *Cytotherapy* 2012; 14(1): 56-60.
- [108] Mazzini L, Ferrero I, Luparello V, *et al.* Mesenchymal stem cell transplantation in amyotrophic lateral sclerosis: A Phase I clinical trial. *Exp Neurol* 2010; 223(1): 229-37.
- [109] Prabhakar S, Marwaha N, Lal V, *et al.* Autologous bone marrow-derived stem cells in amyotrophic lateral sclerosis: a pilot study. *Neurol India* 2012; 60(5): 465-9.
- [110] Karussis D, Karageorgiou C, Vaknin-Dembinsky A, *et al.* Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis. *Arch Neurol* 2010; 67(10): 1187-94.
- [111] Baek W, Kim Y, Koh S, *et al.* Stem cell transplantation into the intraventricular space via an Ommaya reservoir in a patient with amyotrophic lateral sclerosis. *J Neurosurg Sci* 2012; 56(3): 261-3.
- [112] Martinez HR, Gonzalez-Garza MT, Moreno-Cuevas JE, *et al.* Stem-cell transplantation into the frontal motor cortex in amyotrophic lateral sclerosis patients. *Cytotherapy* 2009; 11(1): 26-34.
- [113] Appel SH. Is ALS a systemic disorder? Evidence from muscle mitochondria. *Exp Neurol* 2006; 198(1): 1-3.
- [114] Nicaise C, Mitrecic D, Pochet R. Brain and spinal cord affected by amyotrophic lateral sclerosis induce differential growth factors expression in rat mesenchymal and neural stem cells. *Neuropathol appl Neurobiol* 2011; 37(2): 179-88.
- [115] Koh S-H, Baik W, Noh MY, *et al.* The functional deficiency of bone marrow mesenchymal stromal cells in ALS patients is proportional to disease progression rate. *Exp Neurol* 2012; 233(1): 472-80.
- [116] Nachmany H, Wald S, Abekasis M, Bulvik S, Weil M. Two potential biomarkers identified in mesenchymal stem cells and leuko-

- cytes of patients with sporadic amyotrophic lateral sclerosis. *Dis Markers* 2012; 32(4): 211-20.
- [117] Koh S-H, Huh Y-M, Noh MY, *et al.* β -PIX is critical for transplanted mesenchymal stromal cell migration. *Stem Cells Dev* 2011; 21(11): 1989-99.
- [118] Park H-W, Cho J-S, Park C-K, *et al.* Directed induction of functional motor neuron-like cells from genetically engineered human mesenchymal stem cells. *PLoS One* 2012; 7(4): e35244.
- [119] Feng Y, Zhu M, Dangelmajer S, *et al.* Hypoxia-cultured human adipose-derived mesenchymal stem cells are non-oncogenic and have enhanced viability, motility, and tropism to brain cancer. *Cell Death Dis* 2014; 5(12): e1567.
- [120] Kwon MS, Noh MY, Oh KW, *et al.* The immunomodulatory effects of human mesenchymal stem cells on peripheral blood mononuclear cells in ALS patients. *J Neurochem* 2014; 131(2): 206-18.
- [121] Łukaszewicz-Zajac M, Mroczko B, Słowik A. Matrix metalloproteinases (MMPs) and their tissue inhibitors (TIMPs) in amyotrophic lateral sclerosis (ALS). *J Neural Transm* 2014; 121(11): 1387-97.
- [122] Giunti D, Parodi B, Usai C, *et al.* Mesenchymal stem cells shape microglia effector functions through the release of CX3CL1. *Stem Cells* 2012; 30(9): 2044-53.
- [123] Tripodo G, Chlapanidas T, Perteghella S, *et al.* Mesenchymal Stromal Cells Loading Curcumin-INVITE-Micelles: a Drug Delivery System for Neurodegenerative Diseases. *Colloids Surf B Biointerfaces* 2015; 125: 300-8.
- [124] Wei X, Yang X, Han Z-p, *et al.* Mesenchymal stem cells: a new trend for cell therapy. *Acta Pharmacol Sin* 2013; 34(6): 747-54.
- [125] Mazzini L, Vercelli A, Ferrero I, *et al.* Transplantation of mesenchymal stem cells in ALS. *Prog Brain Res* 2011; 201: 333-59.
- [126] Bocelli-Tyndall C, Zajac P, Di Maggio N, *et al.* Fibroblast growth factor 2 and platelet-derived growth factor, but not platelet lysate, induce proliferation-dependent, functional class II major histocompatibility complex antigen in human mesenchymal stem cells. *Arthritis Rheum* 2010; 62(12): 3815-25.
- [127] Tarte K, Gaillard J, Lataillade J-J, *et al.* Clinical-grade production of human mesenchymal stromal cells: occurrence of aneuploidy without transformation. *Blood* 2010; 115(8): 1549-53.
- [128] Rubio D, Garcia S, Paz MF, *et al.* Molecular characterization of spontaneous mesenchymal stem cell transformation. *PloS one* 2008; 3(1): e1398.
- [129] Wang Y, Huso D, Harrington J, *et al.* Outgrowth of a transformed cell population derived from normal human BM mesenchymal stem cell culture. *Cytherapy* 2005; 7(6): 509-19.
- [130] Tátrai P, Szepesi Á, Matula Z, *et al.* Combined introduction of Bmi-1 and hTERT immortalizes human adipose tissue-derived stromal cells with low risk of transformation. *Biochem Biophys Res Commun* 2012; 422(1): 28-35.

Received: June 29, 2014

Revised: July 30, 2015

Accepted: August 09, 2015